Instructions For Use

MLA-50 Rotor

For Use in the Beckman Coulter
Optima MAX-XP Tabletop Ultracentrifuge

B03896AE
September 2016

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BECKMAN COULTER

MLA-50 Rotor
For Use in the Beckman Coulter Optima MAX-XP Tabletop Ultracentrifuge
B03896AE (September 2016)

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- Outside of the USA and Canada, contact your local Beckman Coulter Representative.

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Revision History

For labeling updates, go to www.beckmancoulter.com and download the latest version of the manual or system help for your instrument.

Issue AD, 07/2016
Updates were made to the following sections: *Specifications*, *Available Tubes for the MLA-50 Rotor*, *Run Times*.

Issue AE, 09/2016
Updates were made to the following sections: *Temperature Limits*.

Note: *Changes that are part of the most recent revision are indicated in text by a bar in the margin of the amended page.*

Safety Notice

Read all product manuals and consult with Beckman Coulter-trained personnel before attempting to use this equipment. Do not attempt to perform any procedure before carefully reading all instructions. Always follow product labeling and manufacturer's recommendations. If in doubt as to how to proceed in any situation, contact your Beckman Coulter Representative.

This safety notice summarizes information basic to the safe use of the rotor described in this manual. The international symbol displayed to the left is a reminder to the user that all safety instructions should be read and understood before operation or maintenance of this equipment is attempted. When you see the symbol on other pages of this publication, pay special attention to the safety information presented. Observance of safety precautions will also help to avoid actions that could damage or adversely affect the performance of the rotor. This rotor was developed, manufactured, and tested for safety and reliability as part of a Beckman Coulter ultracentrifuge/rotor system. Its safety or reliability cannot be assured if used in a centrifuge not of Beckman Coulter's manufacture or in a Beckman Coulter ultracentrifuge that has been modified without Beckman Coulter's approval.

Alerts for Warning, Caution, and Note

⚠ WARNING

WARNING indicates a potentially hazardous situation which, if not avoided, could result in death or serious injury.

⚠ CAUTION

CAUTION indicates a potentially hazardous situation, which, if not avoided, may result in minor or moderate injury. It may also be used to alert against unsafe practices.

NOTE NOTE is used to call attention to notable information that should be followed during installation, use, or servicing of this equipment.

Safety Information for the MLA-50 Rotor

Handle body fluids with care because they can transmit disease. No known test offers complete assurance that such fluids are free of micro-organisms. Some of the most virulent—Hepatitis (B and C) viruses, HIV (I–V), atypical mycobacteria, and certain systemic fungi—further emphasize the need for aerosol protection. Handle other infectious samples according to good laboratory procedures and methods to prevent spread of disease. Because spills may generate aerosols, observe proper safety precautions for aerosol containment. Do not run toxic, pathogenic, or radioactive materials in this rotor without taking appropriate safety precautions. Biosafe containment should be used when Risk Group II materials (as identified in the World Health Organization *Laboratory*

Biosafety Manual) are handled; materials of a higher group require more than one level of protection.

The rotor and accessories are not designed for use with materials capable of developing flammable or explosive vapors. Do not centrifuge such materials in nor handle or store them near the ultracentrifuge.

Although rotor components and accessories made by other manufacturers may fit in the MLA-50 rotor, their safety in this rotor cannot be ascertained by Beckman Coulter. Use of other manufacturers' components or accessories in the MLA-50 rotor may void the rotor warranty and should be prohibited by your laboratory safety officer. Only the components and accessories listed in this publication should be used in this rotor.

Make sure that filled containers are loaded symmetrically into the rotor and that opposing tubes are filled to the same level with liquid of the same density. Make sure that cavities in use have the proper spacers and/or floating spacers inserted before installing the rotor lid.

If disassembly reveals evidence of leakage, you should assume that some fluid escaped the rotor. Apply appropriate decontamination procedures to the centrifuge and accessories.

Never exceed the maximum rated speed of the rotor and labware in use. Refer to the section on *Run Speeds*, and derate the run speed as appropriate.

Do not use sharp tools on the rotor that could cause scratches in the rotor surface. Corrosion begins in scratches and may open fissures in the rotor with continued use.

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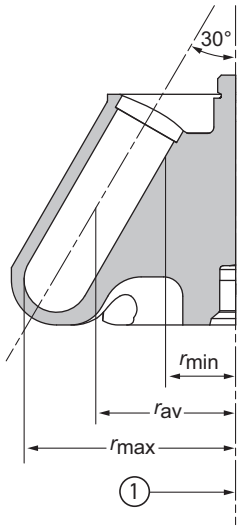
MLA-50 Fixed Angle Rotor

Specifications



1. Axis of Rotation

Maximum speed 50,000 rpm

Density rating at maximum speed 1.2 g/mL

Relative Centrifugal Field[a] at maximum speed

At r_{max} (83.2 mm) $233{,}000 \times g$

At r_{av} (58.4 mm) $164{,}000 \times g$

At r_{min} (33.6 mm) $94{,}100 \times g$

k factor at maximum speed 92[b]

Conditions requiring speed reductions see *Run Speeds*

Number of tube cavities 6

Available tubes see Table 1

Nominal tube dimensions (largest tube) 25 ´ 77 mm

Nominal tube capacity (largest tube) 29.9 mL

Nominal rotor capacity 194.4 mL

Approximate acceleration time to maximum speed (fully loaded) 9 min

Approximate deceleration time from maximum speed (fully loaded 5 min

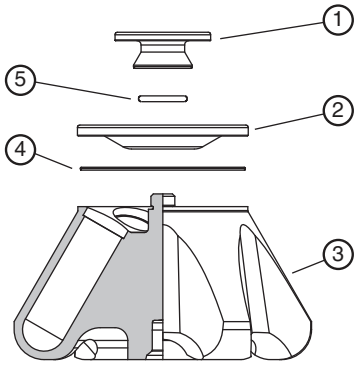
Weight of fully loaded rotor 2.85 kg (6.3 lb)

Rotor and lid material aluminum

a. Relative Centrifugal Field (RCF) is the ratio of the centrifugal acceleration at a specified radius and speed (rw^2) to the standard acceleration of gravity (g) according to the following formula: RCF = rω2/g — where r is the radius in millimeters, ω is the angular velocity in radians per second (2 π RPM /60), and g is the standard acceleration of gravity (9807 mm/s^2). After substitution: RCF = 1.12r (RPM/1000)^2

b. Determined using the 29.9 mL OptiSeal tube.

Description



1. Handle (A96399)
2. Lid (A91766)
3. Rotor Body
4. Large O-ring (A92167)
5. Small O-ring (11521)

This Beckman Coulter rotor has been manufactured in an ISO 9001 or 13485 facility for use with the specified Beckman Coulter ultracentrifuges.

The MLA-50 fixed angle rotor, rated for 50,000 rpm, has a tube angle of 30 degrees from the axis of rotation. The rotor can centrifuge up to six tubes and is used in the Beckman Coulter Optima MAX-XP, tabletop ultracentrifuge.

The rotor, lid, and handle are made of aluminum and anodized to resist corrosion. A rotor retention mechanism on the ultracentrifuge drive hub secures the rotor during the run. Lubricated O-rings made of Buna-N rubber in the rotor lid and handle maintain atmospheric pressure inside the rotor during centrifugation. The tube cavities are numbered to aid in sample identification.

The ultracentrifuge identifies rotor speed during the run by means of a magnetic speed sensor in the instrument chamber and magnets on the bottom of the rotor. This overspeed protection system ensures that the rotor does not exceed its maximum permitted speed.

See the Ultracentrifuge Rotor Warranty at the back of this manual for warranty information.

Preparation and Use

Specific information about the MLA-50 rotor is given here. Information common to this and other rotors is contained in Rotors and Tubes for Tabletop Preparative Ultracentrifuges (publication TLR-IM), which should be used together with this manual for complete rotor and accessory operation. Publication TLR-IM is included in the literature package shipped with the rotor.

NOTE Although rotor components and accessories made by other manufacturers may fit in the MLA-50 rotor, their safety in this rotor cannot be ascertained by Beckman Coulter. Use of other manufacturers' components or accessories in the MLA-50 rotor may void the rotor warranty and should be prohibited by your laboratory safety officer. Only the components and accessories listed in this publication should be used in this rotor.

Prerun Safety Checks

Read the Safety Notice page at the front of this manual before using the rotor.

1 Inspect the rotor assembly (rotor, lid, handle, and O-rings) for damage—the high forces generated in this rotor can cause damaged components to fail.

2 Make sure to use only tubes and accessories listed in Table 1.

3 Check the chemical compatibilities of all materials used.

- Refer to *Chemical Resistances* (publication IN-175), included in the *Rotors and Tubes* CD.

Rotor Preparation

For runs at other than room temperature, refrigerate or warm the rotor beforehand for fast equilibration.

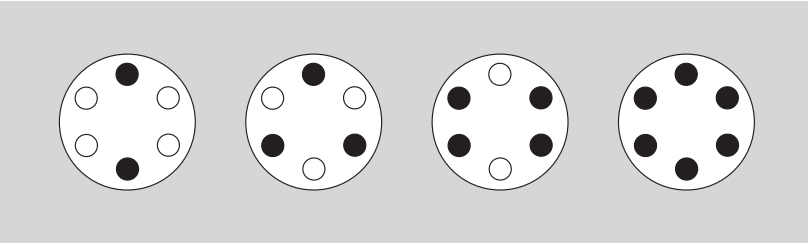
1 Lightly but evenly lubricate metal threads with Spinkote lubricant (306812).

2 Apply a thin film of silicone vacuum grease (335148) to the O-rings in the rotor lid and handle.

3 Load the filled and sealed (if applicable) tubes symmetrically into the rotor (see Table 1 for tube information).

- If fewer than six tubes are being run, they must be arranged symmetrically in the rotor (see Figure 1).
- *Opposing tubes must be filled to the same level with liquid of the same density.*

Figure 1 Arranging Tubes in the Rotor



NOTE Two, three, four, or six tubes can be centrifuged per run if they are arranged in the rotor as shown in Figure 1.

4 Use the required spacers, if necessary (see Table 1), to complete the loading operation.

⚠ CAUTION

Before each run, make sure that the handle is securely tightened.

Operation

1 Use an absorbent towel to wipe off condensation from the rotor.

2 Carefully place the rotor on the drive hub.

3 Refer to the instrument instruction manual for ultracentrifuge operation.

4 For additional operating information, see the following:

- *Run Times*, page 9, for using k factors to adjust run durations.
- *Run Speeds*, page 10, for information about speed limitations.
- *Selecting CsCl Gradients*, page 14, for methods to avoid CsCl precipitation during centrifugation.

Removal and Sample Recovery

⚠ CAUTION

If disassembly reveals evidence of leakage, you should assume that some fluid escaped the rotor. Apply appropriate decontamination procedures to the centrifuge and accessories.

1 Remove the rotor from the ultracentrifuge.

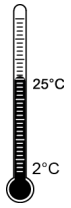
2 Remove the handle by unscrewing it to the left (counterclockwise).

a. Remove the lid.

3 Use a tube removal tool to remove the spacers and tubes.

Tubes and Accessories

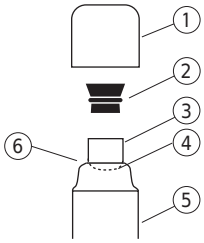
The MLA-50 rotor uses tubes and accessories listed in Table 1. Be sure to use only those items listed, and to observe the maximum speed limits shown. Refer to *Chemical Resistances* for information on the chemical resistances of tube and accessory materials.



Temperature Limits

- Plastic tubes have been centrifuge tested for use at temperatures between 2 and 25°C. For centrifugation at other temperatures, pretest tubes under anticipated run conditions.
- If plastic containers are frozen before use, make sure that they are thawed to at least 2°C prior to centrifugation.

OptiSeal Tubes



1. Spacer
2. Plug
3. Stem
4. Meniscus
5. Tube
6. Base of Stem

OptiSeal tubes come with plastic plugs and can be quickly and easily prepared for use. With the tube spacer in place, the *g* force during centrifugation ensures a tight, reliable seal that protects your samples.

1 Place the tubes in the rack and fill each tube to the base of the stem, leaving no fluid in the stem.

- Overfilling the tube can cause spillage when the plug is inserted or can compromise seal integrity.
- However, too much air can cause excessive tube deformation, disrupting gradients and sample bands.

2 Refer to *Using OptiSeal Tubes* (publication IN-189), included in each box of tubes, for detailed information on the use and care of OptiSeal tubes.

Table 1 Available Tubes for the MLA-50 Rotor[a]

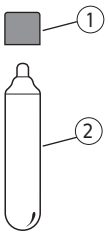
Tube			Required Accessory		Tube Rack	Max Speed/RCF/ *k* factor
Dimensions/Nominal Volume/	**Description**	**Part Number**	**Description**	**Part Number**		
25 × 77 mm 29.9 mL	OptiSeal Polypropylene	361625 (pkg/56)	amber Ultem[b] spacer	361669	348124	50,000 rpm 233,000 × *g* 92
25 × 64 mm 27.0 mL	Quick-Seal Polypropylene	343665 (pkg/50)	Noryl[c] floating spacer	343448	348124	50,000 rpm 233,000 × *g* 79
25 × 64 mm 27.0 mL	Quick-Seal Polypropylene	357346[d] (pkg/50)	Noryl floating spacer	343448	348124	50,000 rpm 233,000 × *g* 79
25 × 38 mm 15.0 mL	Quick-Seal Polypropylene	343664 (pkg/50)	Noryl floating spacer	343448	348124	50,000 rpm 233,000 × *g* 50
25 × 38 mm 15.0 mL	Quick-Seal Polypropylene	357343[d] (pkg/50)	Noryl floating spacer	343448	348124	50,000 rpm 233,000 × *g* 50
25 × 64 mm 27.0 mL	Quick-Seal Ultra-Clear	344323 (pkg/50)	Noryl floating spacer	343448	348124	50,000 rpm 233,000 × *g* 79
25 × 38 mm 15.0 mL	Quick-Seal Ultra-Clear	344324 (pkg/50)	Noryl floating spacer	343448	348124	50,000 rpm 233,000 × *g* 50
16 × 76 mm 13.5 mL	thinwall Polypropylene	326814 (pkg/50)	cap assembly	330860	348122	50,000 rpm 209,000 × *g* 94
			Delrin[e] adapter	303307		
13 × 64 mm 6.5 mL	thinwall Polypropylene	326820 (pkg/50)	cap assembly	346256[f]	348122	50,000 rpm 189,000 × *g* 76
			Delrin adapter	303392		
16 × 76 mm 10.0 mL	thickwall Polypropylene	355640 (pkg/25)	aluminum cap	338907	348123	50,000 rpm 206,000 × *g* 89
			Delrin adapter	303307		
13 × 64 mm 4.0 mL	thickwall Polypropylene	355644 (pkg/25)	adapter	303392	348122	50,000 rpm 187,000 × *g* 52

Table 1 Available Tubes for the MLA-50 Rotor[a] *(Continued)*

Tube			Required Accessory		Tube Rack	Max Speed/RCF/ *k* factor
Dimensions/Nominal Volume/	**Description**	**Part Number**	**Description**	**Part Number**		
16 × 76 mm 10.0 mL	thickwall polycarbonate	355630 (pkg/25)	aluminum cap	338907	348123	50,000 rpm 206,000 × *g* 89
			Delrin adapter	303307		
13 × 64 mm 4.0 mL	thickwall polycarbonate	355645 (pkg/25)	adapter	303392	348122	50,000 rpm 187,000 × *g* 52

a. Use only the items listed here and observe maximum fill volumes and speeds shown.
b. Ultem is a registered trademark of GE Plastics.
c. Noryl is a registered trademark of GE Plastics.
d. To simplify ordering, these g-Max kits include 12 spacers, 50 tubes, and required tools.
e. Delrin is a registered trademark of E.I. Du Pont de Nemours & Company.
f. Cap assembly includes neoprene gasket (344672) and stem (346246).

Quick Seal Tubes



1. *g*-Max Spacer
2. Bell-top Tube

Quick-Seal tubes must be sealed prior to centrifugation. These tubes are heat sealed and do not need caps; however, spacers are required on top of the tubes when they are loaded into the rotor

1 Fill Quick-Seal tubes leaving a *small* bubble of air at the base of the neck.

a. Do not leave a large air space—too much air can cause excessive tube deformation.

2 Refer to *Rotors and Tubes* for detailed information on the use and care of Quick-Seal tubes.

Some of the tubes listed in Table 1 are part of the *g*-Max system. The *g*-Max system uses a combination of small bell-top Quick-Seal tubes and floating spacers (also called *g*-Max spacers). This means that you can run the shorter tubes listed in the table in the MLA-50 rotor without reduction in *g* force. Additional information about the *g*-Max system is available in publication DS-709B.

Thinwall Tubes

Thinwall Polypropylene and Ultra-Clear tubes require caps for tube support.

1 Fill the tubes as full as possible to prevent tube collapse during centrifugation.

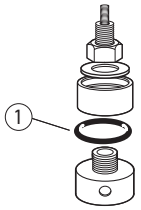
2 If necessary, float mineral oil (or some other low-density, immiscible liquid) on top of the tube contents to fill the tube to its maximum volume.

a. (Do not use an oil overlay in Ultra-Clear tubes.)

Thickwall Tubes

Thickwall Polypropylene and polycarbonate tubes can be run partially filled (at least half filled) with or without caps, but all opposing tubes for a run must be filled to the same level with liquid of the same density. Do not overfill capless tubes; be sure to note the reductions in fill volume and run speed shown in Table 1.

Tube Caps



1. O-Ring or Gasket

Thinwall tubes require caps for tube support during centrifugation; thickwall tubes and bottles can be run with or without caps (speed reduction may be required). Use only the caps listed in Table 1. Refer to *Rotors and Tubes* for the use, assembly, and maintenance of caps.

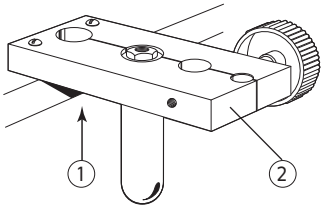
1 Inspect tube caps before use as described in *Rotors and Tubes*.

a. Replace any damaged components.

2 Inspect the O-rings or gaskets in the caps for cracks, nicks, or flattened areas.

a. Be sure that they are dry and free of lubricant during assembly.

3 Hand tighten caps with a hex driver (841883) while the tube is held in the tube-cap vise (305075).



1. Insert tube from bottom **2.** Tube Cap Vise (305075)

Run Times

The *k* factor of the rotor is a measure of the rotor's pelleting efficiency. (Beckman Coulter has calculated the *k* factors for all of its preparative rotors at maximum rated speed and using full tubes.)* The *k* factor is calculated from the formula

$$k = \frac{\ln(r_{max}/r_{min})}{\omega^2} \times \frac{10^{13}}{3600} \quad \text{EQ 1}$$

where ω is the angular velocity of the rotor in radians per second ($\omega = 0.105 \times \text{rpm}$), r_{max} is the maximum radius, and r_{min} is the minimum radius.

After substitution:

$$k = \frac{(2.533 \times 10^{11})\ln(r_{max}/r_{min})}{\text{rpm}^2} \quad \text{EQ 2}$$

Use the *k* factor in the following equation to estimate the run time *t* (in hours) required to pellet particles of known sedimentation coefficient *s* (in Svedberg units, *S*).

$$t = \frac{k}{s} \quad \text{EQ 3}$$

Run times can be estimated for centrifugation at less than maximum speed by adjusting the *k* factor as follows:

$$k_{adj} = k\left(\frac{50{,}000}{\text{actual run speed}}\right)^2 \quad \text{EQ 4}$$

* For the MLA-50 rotor, the k factor was calculated using the maximum volume allowed in a 29.9 mL OptiSeal tube (361625).

Run times can also be estimated from data established in prior experiments if the *k* factor of the previous rotor is known. For any two rotors, a and b:

$$\frac{t_a}{t_b} = \frac{k_a}{k_b} \qquad \textbf{EQ 5}$$

Run Speeds

The centrifugal force at a given radius in a rotor is a function of speed. Comparisons of forces between different rotors are made by comparing the rotors' relative centrifugal fields (RCF). When rotational speed is adjusted so that identical samples are subjected to the same RCF in two different rotors, the samples are subjected to the same force. The RCF at a number of rotor speeds is provided in Table 2.

Speeds must be reduced under the following circumstances:

1. If nonprecipitating solutions more dense than 1.2 g/mL are centrifuged, the maximum allowable run speed must be reduced according to the following equation:

   $$\text{reduced maximum speed} = (50{,}000 \text{ rpm}) \sqrt{\frac{1.2 \text{ g/mL}}{\rho}} \qquad \textbf{EQ 6}$$

   where ρ is the density of the tube contents. This speed reduction will protect the rotor from excessive stresses due to the added tube load. *Note, however, that the use of this formula may still produce maximum speed values that are higher than the limitations imposed by the use of certain tubes or adapters.* In such cases, use the lower of the two values.
2. *Further speed limits must be imposed* when CsCl or other self-forming-gradient salts are centrifuged, as equation (6) does not predict concentration limits/speeds that are required to avoid precipitation of salt crystals. Precipitation during centrifugation would alter the density distribution of CsCl and this would change the position of the sample bands. Figure 2 and Figure 3, together with the description and examples below, show how to reduce run speeds when using CsCl gradients.

Table 2 Relative Centrifugal Fields for the MLA-50 Rotor[a]

Rotor Speed (rpm)	Relative Centrifugal Field (× *g*)			*k* Factor[b]
	At rmax (83.2 mm)	At rav (58.4 mm)	At rmin (33.6 mm)	
50,000	233,000	164,000	94,100	92
45,000	189,000	132,000	76,200	113
40,000	149,000	105,000	60,200	144
35,000	114,000	80,100	46,100	188
30,000	83,900	58,900	33,900	255
25,000	58,200	40,900	23,500	368
20,000	37,300	26,200	15,000	574
15,000	21,000	14,700	8,470	1021
10,000	9,320	6,540	3,760	2297
5,000	2,330	1,640	940	9187

a. Entries in this table are calculated from the formula RCF = 1.12r (RPM/1000)2 and then rounded to three significant digits.

b. Calculated for all Beckman Coulter preparative rotors as a measure of the rotor's relative efficiency in pelleting sample in water at 20°C.

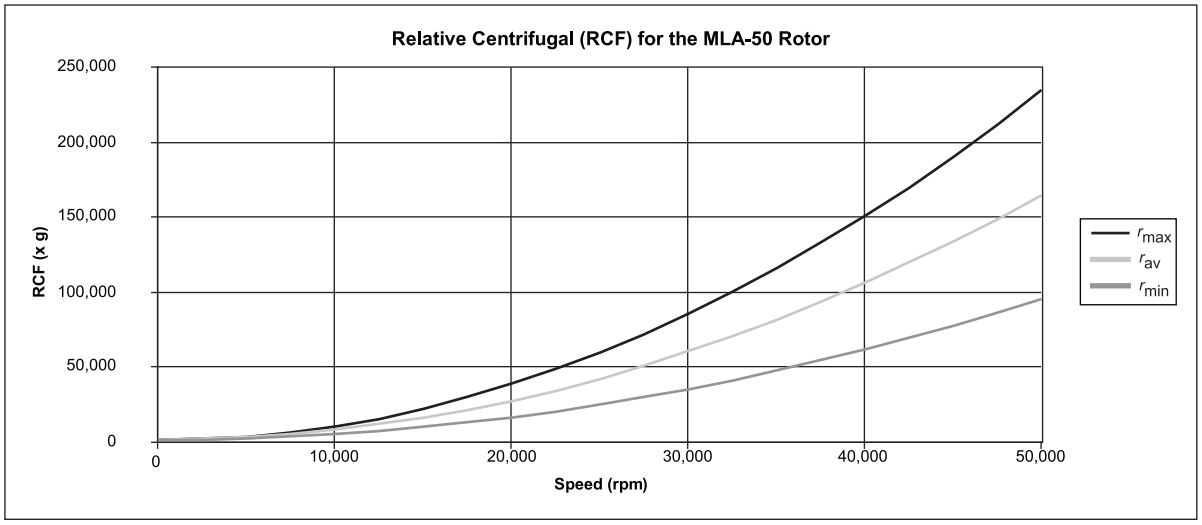
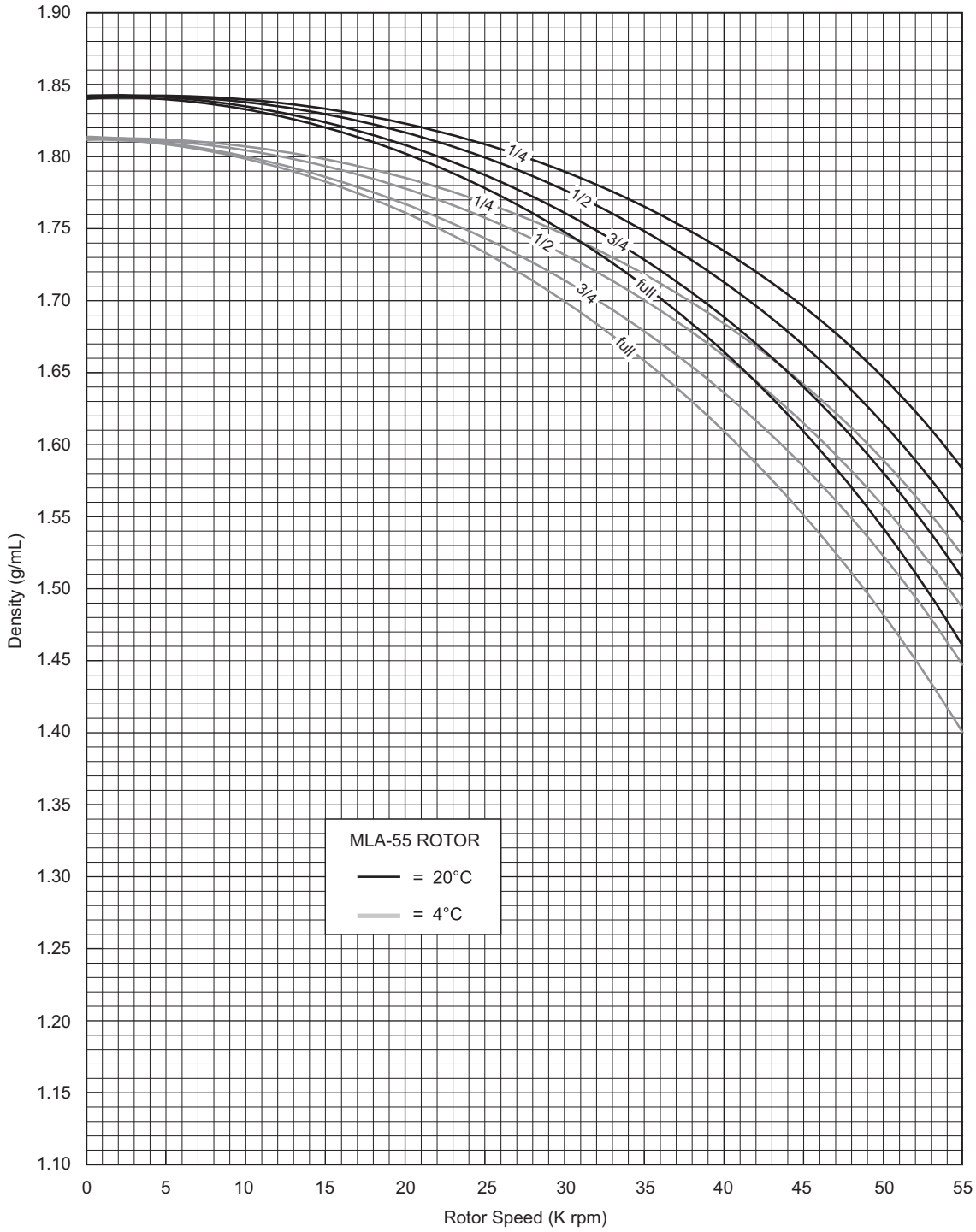
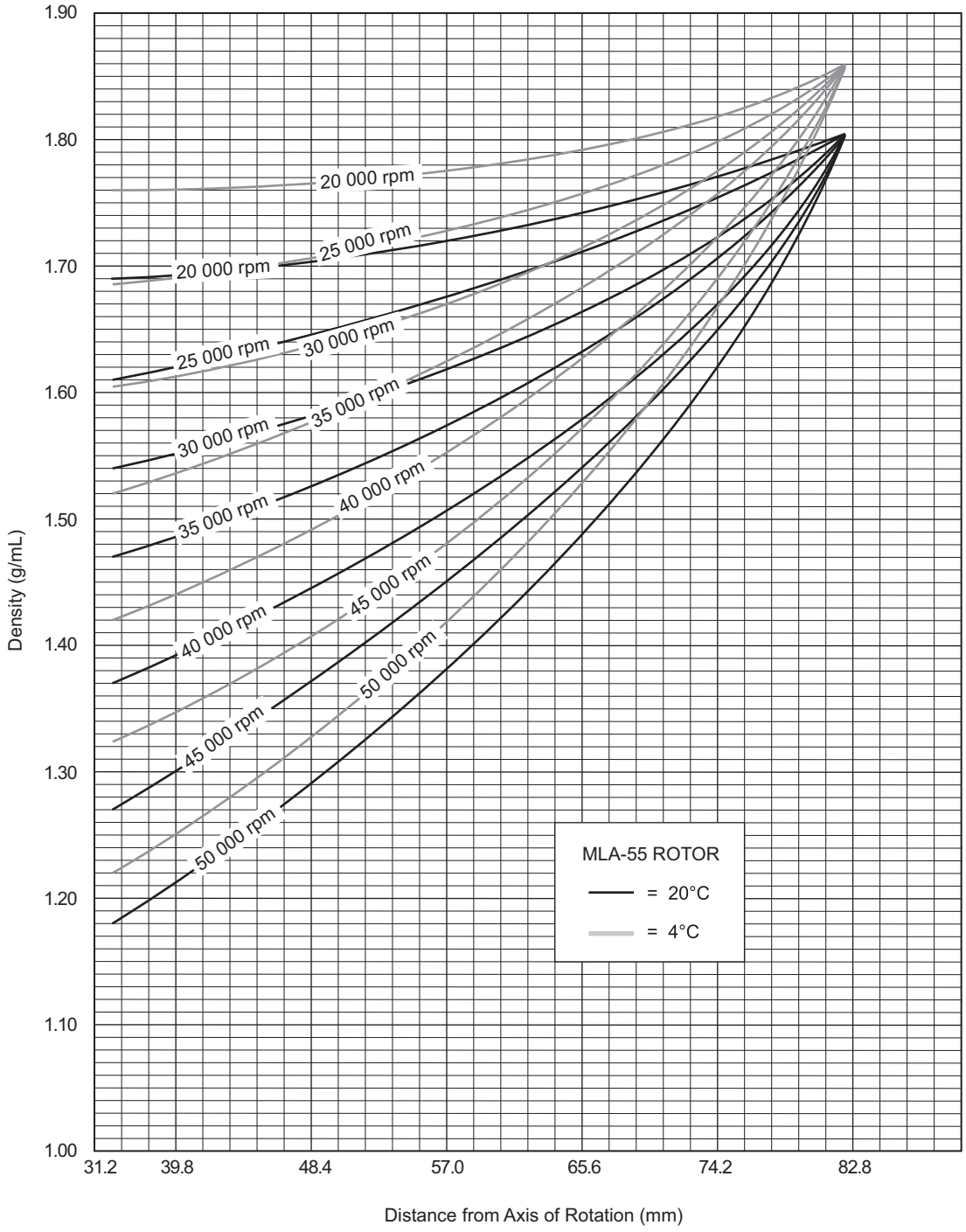


Figure 2 Precipitation Curves for the MLA-50 Rotor*



* Using combinations of rotor speeds and homogeneous CsCl solution densities that intersect on or below these curves ensures that CsCl will not precipitate during centrifugation.

Figure 3 CsCl Gradients at Equilibrium for the MLA-50 Rotor*



* Centrifugation of homogeneous CsCl solutions at the maximum allowable speeds (from Figure 2) results in gradients presented here.

Selecting CsCl Gradients

Precipitation during centrifugation would alter density distribution, and this would change the position of the sample bands. Curves in Figure 2 and Figure 3 are provided up to the maximum rated speed of the rotor.

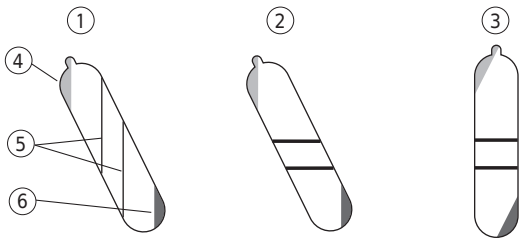
NOTE The curves in Figure 2 and Figure 3 are for solutions of CsCl salt dissolved in distilled water only. If other salts are present in significant concentrations, the overall CsCl concentration may need to be reduced.

Rotor speed is used to control the slope of a CsCl density gradient, and must be limited so that CsCl precipitation is avoided. Speed and density combinations that intersect on or below the curves in Figure 2 ensure that CsCl will not precipitate during centrifugation in the MLA-50 rotor. Curves are provided at two temperatures: 20°C (black curves) and 4°C (gray curves).

The reference curves in Figure 3 show gradient distribution at equilibrium. Each curve in Figure 3 is within the density limits allowed for the MLA-50 rotor: each curve was generated for a single run speed using the maximum allowable homogeneous CsCl densities (one for each fill level) that avoid precipitation at that speed. (The gradients in Figure 3 can be generated from step or linear gradients, or from homogeneous solutions. But the total amount of CsCl in solution must be equivalent to a homogeneous solution corresponding to the concentrations specified in Figure 2.) Figure 3 can also be used to approximate the banding positions of sample particles. Gradient curves not shown in Figure 3 can be interpolated.

Typical Examples for Determining CsCl Run Parameters

Example A:
A separation that is done frequently is the banding of plasmic DNA in cesium chloride with ethidium bromide. The starting density of the CsCl solution is 1.55 g/mL. In this separation the covalently closed, circular plasmid bands at a density of 1.57 g/mL, while the nicked and linear species band at 1.53 g/mL. At 20°C, where will particles band?



1. At Speed
2. At Rest
3. Upright
4. Floating Components
5. Bands
6. Pelleted Material

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1 In Figure 2, find the curve that corresponds to the desired run temperature (20°C) and fill volume (full).

- The maximum allowable rotor speed is determined from the point where this curve intersects the homogeneous CsCl density (44,000 rpm)

2 In Figure 3, sketch in a horizontal line corresponding to each particle's buoyant density.

3 Mark the point in the figure where each particle density intersects the curve corresponding to the selected run speed and temperature.

- Particles will band at these locations across the tube diameter at equilibrium during centrifugation.

In this example, particles will band about 61.0 and 57.9 mm from the axis of rotation, about 2.0 mm of center-of-band to center-of-band separation at the rotor's 30-degree tube angle. When the tube is removed from the rotor and held upright (vertical and stationary), there will be about 3.7 mm of centerband-to-centerband separation. This interband distance, d_{up}, can be calculated from the formula:

$$d_{up} = \frac{d_\theta}{\cos\theta} \qquad \text{EQ 7}$$

where d_q is the interband distance when the tube is held at an angle, θ, in the rotor.

Example B:
Knowing particle buoyant densities (such as 1.59 and 1.54 g/mL), how do you achieve good separation?

1 In Figure 3, sketch in a horizontal line corresponding to each particle's buoyant density.

2 Select the curve at the desired temperature (20°C) that gives the best particle separation.

3 Note the run speed along the selected curve (35,000 rpm).

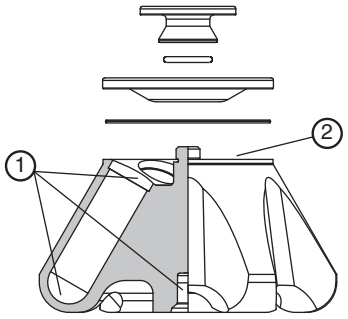
4 From Figure 2, select the maximum homogeneous CsCl density (in this case, 1.66 g/mL) that corresponds to the temperature and run speed established above.

- These parameters will provide the particle-banding pattern selected in Step 2.

In this example, particles will band at about 46.6 and 36.3 mm from the axis of rotation (about 10.3 mm apart). When the tube is held upright there will be about 11.9 mm of center-of-band to center-of-band separation.

Care and Maintenance

Maintenance



1. Check for Corrosion
2. Rotor Body

NOTE Do not use sharp tools on the rotor that could cause scratches in the rotor surface. Corrosion begins in scratches and may open fissures in the rotor with continued use.

1 Periodically (at least monthly) inspect the rotor, specially inside cavities, for rough spots or pitting, white powder deposits (which may be aluminum oxide), or heavy discoloration.

- **a.** If any of these signs are evident, do not run the rotor.
- **b.** Contact your Beckman Coulter representative for information about the Field Rotor Inspection Program and the rotor repair center.

2 Regularly lubricate the metal threads in the rotor with a thin, even coat of Spinkote lubricant (306812).

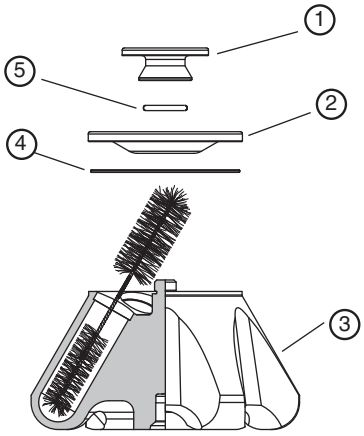
- Failure to keep these threads lubricated can result in damaged threads.

3 Regularly apply silicone vacuum grease to the O-rings.

- **a.** Replace the O-rings about twice a year or whenever worn or damaged.

Refer to *Chemical Resistances* for the chemical resistances of rotor and accessory materials. Your Beckman Coulter representative provides contact with the Field Rotor Inspection Program and the rotor repair center.

Cleaning

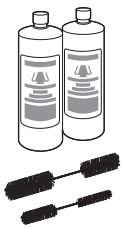


1. Lid Handle
2. Lid
3. Rotor Body
4. Large O-ring (A92167)
5. Small O-ring (11521)

Wash the rotor and rotor components immediately if salts or other corrosive materials are used or if spillage has occurred. Do not allow corrosive materials to dry on the rotor.

Under normal use, wash the rotor frequently (at least weekly) to prevent buildup of residues.

1 Remove the O-rings before washing.



2 Wash the rotor and lid in a mild detergent, such as Beckman Solution 555 (339555), that won't damage the rotor.

- The Rotor Cleaning Kit contains two plastic-coated brushes and two quarts of Solution 555 for use with rotors and accessories.

a. Dilute the detergent 10 to 1 with water.

NOTE Do not wash rotor components in a dishwasher. Do not soak in detergent solution for long periods, such as overnight.

3 Rinse the cleaned rotor and components with distilled water.

4 Air-dry the rotor and lid upside down.

a. *Do not use acetone to dry the rotor.*

5 Apply a thin, even coat of silicone vacuum grease to the O-rings before replacing them in the grooves in the lid and handle.

6 Clean metal threads as necessary (at least every 6 months).

a. Use a brush and concentrated Solution 555.

b. Rinse and dry thoroughly, then lubricate lightly but evenly with Spinkote to coat all threads.

Periodically remove the O-rings and wipe clean as necessary. Clean the O-ring grooves with a cotton-tipped swab. Reapply a light film of silicone vacuum grease.

Decontamination

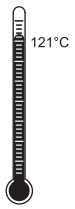
If the rotor (and/or accessories) becomes contaminated with radioactive material, it should be decontaminated using a solution that will not damage the anodized surfaces. Beckman Coulter has tested a number of solutions and found two that do not harm anodized aluminum: RadCon Surface Spray or IsoClean Solution (for soaking),* and Radiacwash.†

NOTE IsoClean can cause fading of colored anodized surfaces. Use it only when necessary and remove it promptly from surfaces.

While Beckman Coulter has tested these methods and found that they do not damage components, no guarantee of decontamination is expressed or implied. Consult your laboratory safety officer regarding the proper decontamination methods to use.

If the rotor or other components are contaminated with toxic or pathogenic materials, follow appropriate decontamination procedures as outlined by your laboratory safety officer.

Sterilization and Disinfection



- The rotor and all rotor components can be autoclaved at 121°C for up to an hour. Remove the O-rings from the handle and lid and place the rotor, handle, and lid in the autoclave upside down.
- Ethanol (70%)‡ or hydrogen peroxide (6%) may be used on all rotor components, including those made of plastic. Bleach (sodium hypochlorite) may be used, but may cause discoloration of anodized surfaces. Use the minimum immersion time for each solution, per laboratory standards.

* In U.S., contact Nuclear Associates (New York); in Eastern Europe and Commonwealth States, contact Victoreen GmbH (Munich); in South Pacific, contact Gammasonics Pty. Ltd. (Australia); in Japan, contact Toyo Medic Co. Ltd. (Tokyo).

† In U.S., contact Biodex Medical Systems (Shirley, New York); internationally, contact the U.S. office to find the dealer closest to you.

‡ Flammability hazard. Do not use in or near operating ultracentrifuges.

While Beckman Coulter has tested these methods and found that they do not damage the rotor or components, no guarantee of sterility or disinfection is expressed or implied. When sterilization or disinfection is a concern, consult your laboratory safety officer regarding proper methods to use.

Storage

When it is not in use, store the rotor in a dry environment (not in the instrument) with the lid removed to allow air circulation so moisture will not collect in the tube cavities.

Returning a Rotor

Before returning a rotor or accessory for any reason, prior permission must be obtained from Beckman Coulter, Inc. This form may be obtained from your local Beckman Coulter sales office. The form, entitled *Returned Material Authorization* (RMA) for United States returns or *Returned Goods Authorization* (RGA) for international returns, should contain the following information:

- rotor type and serial number,
- history of use (approximate frequency of use),
- reason for the return,
- original purchase order number, billing number, and shipping number, if possible,
- name and email address of the person to be notified upon receipt of the rotor or accessory at the factory,
- name and email address of the person to be notified about repair costs, etc.

To protect our personnel, it is the customer's responsibility to ensure that all parts are free from pathogens and/or radioactivity. Sterilization and decontamination must be done before returning the parts. Smaller items (such as tubes, bottles, etc.) should be enclosed in a sealed plastic bag.

All parts must be accompanied by a note, plainly visible on the outside of the box or bag, stating that they are safe to handle and that they are not contaminated with pathogens or radioactivity. ***Failure to attach this notification will result in return or disposal of the items without review of the reported problem.***

Use the address label printed on the RMA/RGA form when mailing the rotor and/or accessories.

Customers located outside the United States should contact their local Beckman Coulter office.

Supply List

NOTE Publications referenced in this manual can be obtained at www.beckmancoulter.com, by calling Beckman Coulter at 1-800-742-2345 in the United States, or by contacting your local Beckman Coulter office.

See the Beckman Coulter *Ultracentrifuge Rotors, Tubes & Accessories* catalog (BR-8101, available at www.beckmancoulter.com) or contact Beckman Coulter Sales (1-800-742-2345 in the United States)

for detailed information on ordering parts and supplies. For your convenience, a partial list is given below.

Replacement Rotor Parts

MLA-50 rotor assembly	A91774
Handle	A96399
Lid	A91766
Handle O-ring (small)	11521
Lid O-ring (large)	A92167

Other

NOTE For MSDS information, go to the Beckman Coulter website at www.beckmancoulter.com.

Tubes, bottles, and accessories	see Table 1
OptiSeal tube rack assembly	361646
Quick-Seal Cordless Tube Topper kit, 60 Hz	358312
Quick-Seal Cordless Tube Topper kit, 50 Hz (Europe)	358313
Quick-Seal Cordless Tube Topper kit, 50 Hz (Great Britain)	358314
Quick-Seal Cordless Tube Topper kit, 50 Hz (Australia)	358315
Tube Topper rack	348124
Tool kit for aluminum caps includes:	331202
Torque wrench	858121
Socket adapter	858122
Socket for 11-mm hex nuts	870432
Socket for 19-mm hex nuts	858123
Tube removal tool	301875
Hex driver (for 11-mm cap nuts)	841883
Hex driver (for 8-mm cap nuts)	841884
Floating spacer removal tool	338765
Tube-cap vise	305075
Tube removal tool (Quick-Seal and OptiSeal tubes)	361668
Spinkote lubricant (2 oz)	306812
Silicone vacuum grease (1 oz)	335148
Rotor Cleaning Kit	339558
Beckman Solution 555 (1 qt)	339555
Rotor cleaning brush	339379

Beckman Coulter, Inc.
Ultracentrifuge Rotor Warranty

All Beckman Coulter ultracentrifuge Fixed Angle, Vertical Tube, Near Vertical Tube, Swinging Bucket, and Airfuge rotors are warranted against defects in materials or workmanship for the time periods indicated below, subject to the Warranty Conditions stated below.

Preparative Ultracentrifuge Rotors	5 years — No Proration
Analytical Ultracentrifuge Rotors	5 years — No Proration
ML and TL Series Ultracentrifuge Rotors	5 years — No Proration
Airfuge Ultracentrifuge Rotors	1 year — No Proration

For Zonal, Continuous Flow, Component Test, and Rock Core Ultracentrifuge Rotors, see separate warranty.

Warranty Conditions (as applicable)

1. This warranty is valid for the time periods indicated above from the date of shipment to the original Buyer by Beckman Coulter or an authorized Beckman Coulter representative.
2. This warranty extends only to the original Buyer and may not be assigned or extended to a third person without written consent of Beckman Coulter.
3. This warranty covers the Beckman Coulter Centrifuge Systems only (including but not limited to the centrifuge, rotor, and accessories) and Beckman Coulter shall not be liable for damage to or loss of the user's sample, non-Beckman Coulter tubes, adapters, or other rotor contents.
4. This warranty is void if the Beckman Coulter Centrifuge System is determined by Beckman Coulter to have been operated or maintained in a manner contrary to the instructions in the operator's manual(s) for the Beckman Coulter Centrifuge System components in use. This includes but is not limited to operator misuse, abuse, or negligence regarding indicated maintenance procedures, centrifuge and rotor classification requirements, proper speed reduction for the high density of certain fluids, tubes, and tube caps, speed reduction for precipitating gradient materials, and speed reduction for high-temperature operation.
5. Rotor bucket sets purchased concurrently with or subsequent to the purchase of a Swinging Bucket Rotor are warranted only for a term co-extensive with that of the rotor for which the bucket sets are purchased.
6. This warranty does not cover the failure of a Beckman Coulter rotor in a centrifuge not of Beckman Coulter manufacture, or if the rotor is used in a Beckman Coulter centrifuge that has been modified without the written permission of Beckman Coulter, or is used with carriers, buckets, belts, or other devices not of Beckman Coulter manufacture.
7. Rotor parts subject to wear, including but not limited to rotor O-rings, VTi, NVT, TLV, MLN, and TLN rotor tube cavity plugs and gaskets, tubing, tools, optical overspeed disks, bearings, seals, and lubrication are excluded from this warranty and should be frequently inspected and replaced if they become worn or damaged.
8. Keeping a rotor log is not mandatory, but may be desirable for maintenance of good laboratory practices.

Repair and Replacement Policies

1. If a Beckman Coulter rotor is determined by Beckman Coulter to be defective, Beckman Coulter will repair or replace it, subject to the Warranty Conditions. A replacement rotor will be warranted for the time remaining on the original rotor's warranty.
2. If a Beckman Coulter centrifuge is damaged due to a failure of a rotor covered by this warranty, Beckman Coulter will supply free of charge (i) all centrifuge parts required for repair (except the drive unit, which will be replaced at the then current price less a credit determined by the total number of revolutions or

years completed, provided that such a unit was manufactured or rebuilt by Beckman Coulter), and (ii) if the centrifuge is currently covered by a Beckman Coulter warranty or Full Service Agreement, all labor necessary for repair of the centrifuge.

3. If a Beckman Coulter rotor covered by this warranty is damaged due to a malfunction of a Beckman Coulter ultracentrifuge covered by an Ultracentrifuge System Service Agreement, Beckman Coulter will repair or replace the rotor free of charge.
4. If a Beckman Coulter rotor covered by this warranty is damaged due to a failure of a Beckman Coulter tube, bottle, tube cap, spacer, or adapter, covered under the Conditions of this Warranty, Beckman Coulter will repair or replace the rotor and repair the instrument as per the conditions in policy point (2) above, and the replacement policy.
5. Damage to a Beckman Coulter rotor or instrument due to the failure or malfunction of a non-Beckman Coulter tube, bottle, tube cap, spacer, or adapter is not covered under this warranty, although Beckman Coulter will assist in seeking compensation under the manufacturer's warranty.

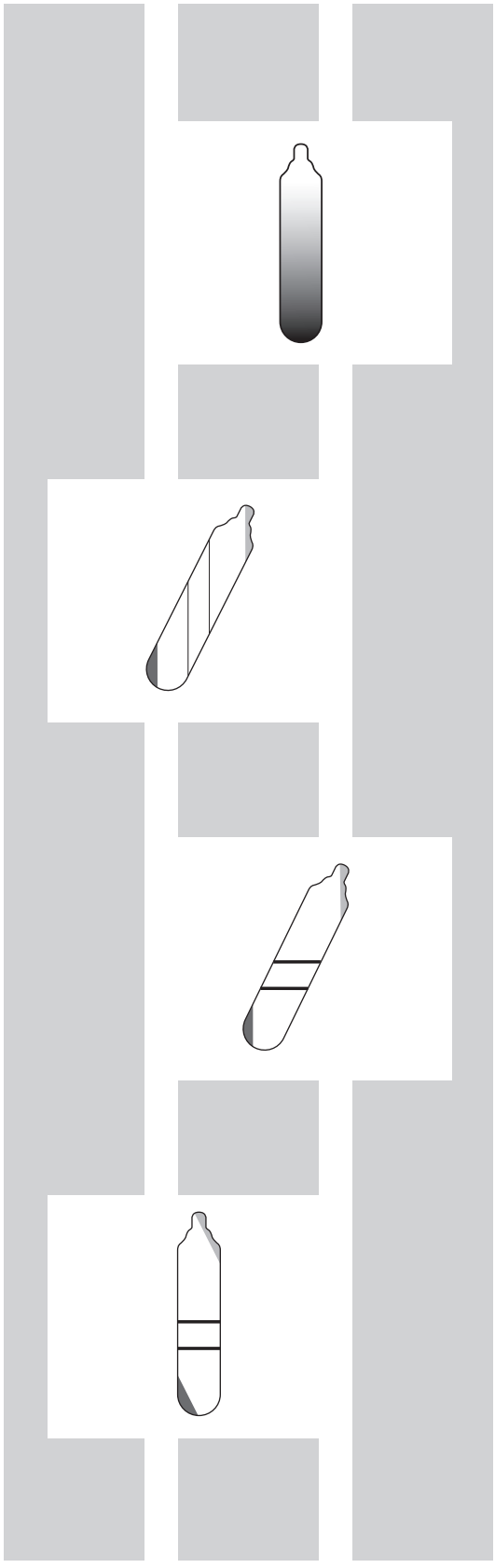
Disclaimer

IT IS EXPRESSLY AGREED THAT THE ABOVE WARRANTY SHALL BE IN LIEU OF ALL WARRANTIES OF FITNESS AND OF THE WARRANTY OF MERCHANTABILITY AND BECKMAN COULTER, INC. SHALL HAVE NO LIABILITY FOR SPECIAL OR CONSEQUENTIAL DAMAGES OF ANY KIND WHATSOEVER ARISING OUT OF THE MANUFACTURE, USE, SALE, HANDLING, REPAIR, MAINTENANCE, OR REPLACEMENT OF THE PRODUCT.

Factory Rotor Inspection Service

Beckman Coulter, Inc., will provide free mechanical and metallurgical inspection in Indianapolis, Indiana, USA, of any Beckman Coulter rotor at the request of the user. (Shipping charges to Beckman Coulter are the responsibility of the user.) Rotors will be inspected in the user's laboratory if the centrifuge in which they are used is covered by an appropriate Beckman Coulter Service Agreement. Contact your local Beckman Coulter office for details of service coverage or cost.

Before shipping, contact the nearest Beckman Coulter Sales and Service office and request a Returned Goods Authorization (RGA) form and packaging instructions. Please include the complete rotor assembly, with buckets, lid, handle, tube cavity caps, etc. A SIGNED STATEMENT THAT THE ROTOR AND ACCESSORIES ARE NON-RADIOACTIVE, NON-PATHOGENIC, NON-TOXIC, AND OTHERWISE SAFE TO SHIP AND HANDLE IS REQUIRED.



Related Documents

Optima MAX-XP Ultracentrifuge (393552)

- Description
- Preinstallation Requirements
- Operation
- Troubleshooting
- Maintenance

Available in hard copy or electronic pdf by request.

Rotors and Tubes for Tabletop Preparative Ultracentrifuges (TLR-IM)

- Rotors
- Tubes and Accessories
- Using Tube and Accessories
- Using Rotors
- Care and Maintenance
- Chemical Resistances for Beckman Coulter Centrifugation Products
- The Use of Cesium Chloride Curves
- Gradient Materials
- Glossary of Terms
- References

Available in hard copy or electronic pdf by request.

Rotors and Tubes CD (369668)

- Rotors and Tubes for Tabletop Preparative Ultracentrifuges
- Rotors and Tubes for J2, J6, Avanti J Series Centrifuges
- Rotors and Tubes for Preparative Ultracentrifuges
- Rotor Safety Bulletin
- Chemical Resistances for Beckman Coulter Centrifugation Products

Additional References

- Chemical Resistances for Beckman Coulter Centrifugation Products (IN-175)
- Beckman Coulter Ultracentrifuge Rotors, Tubes & Accessories catalog (BR-8101)
- Using OptiSeal Tubes (IN-189)

Available in hard copy or electronic pdf by request.

Data Sheets

- *g*-Max System: Short Pathlengths in High Force Fields (DS-709B)

Available at www.beckmancoulter.com

www.beckmancoulter.com

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